Fertility of Holstein Dairy Heifers after Synchronization of Ovulation and Timed AI or AI after Removed Tail Chalk

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ABSTRACT

Nonlactating Holstein dairy heifers ($n = 352$) 13 mo of age were managed using a 42-d artificial insemination (AI) breeding period in which they received AI after removed tail chalk evaluated once daily. At AI breeding period onset (d 0), heifers were randomly assigned to receive synchronization of ovulation ($100 \mu\text{g}$ of GnRH, d 0; 25 mg of $\text{PGF}_{2\alpha}$, d 6; $100 \mu\text{g}$ of GnRH, d 8) and timed AI (TAI; d 8) and AI after removed tail chalk for the entire AI breeding period (GPG; $n = 175$), or AI after removed tail chalk for the entire AI breeding period (TC; $n = 177$). As expected, 17.7% (31/175) of GPG heifers received AI after removed tail chalk before scheduled TAI. Pregnancy rate per artificial insemination (PR/AI) at ~30 d after first AI tended to be greater for TC (46.5%) than for GPG (38.3%) heifers. No treatment $\times$ inseminator interaction was detected; however, overall PR/AI was low for heifers in both treatments due to variation among the 3 inseminators (24.8, 30.0, and 58.0%). Pregnancy loss from ~30 to ~75 d after first AI was 10% and did not differ between treatments. Based on survival analysis, days to first AI was greater for TC than for GPG heifers, whereas days to pregnancy across the 42-d AI breeding period did not differ between treatments. Overall, 81.2% of GPG heifers receiving TAI synchronized luteal regression and ovulated within 48 h after the second GnRH injection. We conclude that this synchronization protocol can yield acceptable fertility in dairy heifers if AI to estrus is conducted between treatment with GnRH and $\text{PGF}_{2\alpha}$ and AI efficiency is optimized.

(Key words: dairy heifer, synchronization of ovulation, timed artificial insemination)

Abbreviation key: PR/AI = pregnancy rate per artificial insemination, P_4 = progesterone, TAI = timed artificial insemination, WHI = wither height index.

INTRODUCTION

Many estrus synchronization protocols have been developed for cattle using progestogens, $\text{PGF}_{2\alpha}$ (or $\text{PGF}_{2\alpha}$ analogs), and estrogens alone or in various combinations to control the physiology of the reproductive cycle and synchronize estrus (Odde, 1990). The primary reason for synchronizing estrus in dairy heifers is to facilitate use of AI (Xu and Burton, 1999). The effectiveness of current estrus synchronization strategies for heifers is limited because dairy producers must rely on visual estrus detection, which is inefficient on most farms, to accurately time AI. In support of this notion, results from a nationwide survey of US dairy producers identified “inadequate heat detection for AI” and “lack of time to supervise AI” as 2 important reasons for not using AI to breed dairy heifers (Erven and Arbaugh, 1987).

Ovsynch was the first protocol developed to successfully synchronize ovulation in lactating dairy cows, thereby allowing for timed artificial insemination (TAI; Pursley et al., 1995). Unfortunately, dairy heifers respond poorly to Ovsynch and TAI, yielding conception rates 20 to 40% lower than heifers receiving AI to a detected estrus (Schmitt et al., 1996; Pursley et al., 1997). Some studies have assessed the effectiveness of synchronizing ovulation using GnRH and $\text{PGF}_{2\alpha}$ followed by TAI in dairy heifers, but results are inconsistent due to low numbers of heifers (Moreira et al., 2000a) or an unknown synchronization response (Pursley et al., 1997). Moreover, previous experiments have tested TAI protocols in dairy heifers using a 7-d interval between GnRH and $\text{PGF}_{2\alpha}$ (Pursley et al., 1997; Moreira et al., 2000a; Lucy et al., 2001).

The objective of this study was to compare the effectiveness of a synchronization protocol using GnRH and $\text{PGF}_{2\alpha}$ followed by TAI to a standard reproductive management system using AI after removed tail chalk in a commercial dairy heifer growing operation in Wisconsin. Our hypothesis was that synchronization of ovulation in dairy heifers using GnRH and $\text{PGF}_{2\alpha}$ followed by TAI would result in improved reproductive performance compared with herd mates managed using AI after removed tail chalk, the standard reproductive management system in place on this farm. Secondary objectives were to assess ovarian responses to the GPG protocol

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and fertility and embryonic losses in dairy heifers under field conditions.

MATERIALS AND METHODS

Facilities and Management of Heifers

This field trial was conducted on a custom dairy heifer growing operation located in central Wisconsin comprising approximately 4500 Holstein dairy heifers. Heifers from 11 different farms arrived at the custom heifer growing operation at around 5 mo of age and were sorted by age to different pens and managed until 13 mo of age in open dry lots throughout the year. Every 10 to 14 d, a group of 80 to 100 heifers within 2 wk of turning 13 mo of age was moved to one of 4 AI breeding pens, each with 100 self-locking head gates. Heifers were fed a TMR based on corn silage, alfalfa haylage, potatoes, and mineral mix once daily between 0630 and 1100 h throughout the AI breeding period. Artificial insemination was conducted for a 42-d period based on removed tail chalk, which was applied and evaluated daily during the morning feeding period after heifers were restrained in the self-locking head gates. After this 42-d AI breeding period, heifers were moved to a dry lot with 2 natural service bulls for about 2 mo. For each group of heifers, pregnancy status was evaluated via palpation per rectum by the herd veterinarian around 60 d after leaving the AI breeding pen to confirm all pregnancies greater than 60 d (AI pregnancies) based on breeding dates and stage of gestation estimated by the veterinarian at the pregnancy diagnosis. Any pregnancies estimated to be less than 60 d were attributed to the natural service bulls. Pregnant heifers stayed in dry lots with natural service bulls and were returned to their farm of origin 6 to 8 wk before their expected calving date. Heifers failing to conceive were culled based on individual owner decision after arrival at their herd of origin.

Treatments

This experiment was conducted in 4 replicates ($n = \sim 88$ heifers/replicate) initiated from May 21 to August 13, 2002. At AI breeding period onset (d 0), heifers were randomly assigned to receive either synchronization of ovulation using GnRH (Cystorelin; Merial Ltd; Duluth, GA) and PGF_{2 α} (Lutalyse; Pharmacia Animal Health, Kalamazoo, MI) and TAI (100 μ g of GnRH, d 0; 25 mg of PGF_{2 α} , d 6; 100 μ g of GnRH+TAI, d 8) followed by AI after removed tail chalk for the remainder of the AI breeding period (GPG; $n = 175$), or AI after removed tail chalk for the entire AI breeding period (TC; $n = 177$). All GPG heifers that did not receive AI to removed tail chalk during the protocol received TAI immediately

after the second GnRH injection (e.g., Cosynch), which was administered 48 h after treatment with PGF_{2 α} . Any GPG heifer detected in estrus before scheduled TAI on d 8 received AI after removed tail chalk, and the synchronization protocol was discontinued. Artificial insemination was performed by 3 herd personnel (the herd manager and 2 full-time employees). A maximum of 39 TAI were performed for GPG heifers on d 8 of the experiment for each replicate.

Blood Sampling and Radioimmunoassay

Blood samples were collected from GPG heifers via venipuncture of the median caudal vein or artery near the base of the tail just before administration of each injection and on the day of AI after removed tail chalk from all heifers regardless of treatment during the first 8 d of the experiment. Blood samples were allowed to clot for 24 h at 4°C, centrifuged (3000 rpm for 15 min), and serum was harvested and stored at -20°C until assayed for progesterone (P₄) concentration by radioimmunoassay (Coat-a-Count Progesterone, Diagnostic Products Corporation, Los Angeles, CA). Inter- and intraassay coefficients of variation were 5.7 and 6.9%, respectively.

Progesterone Classes

All GPG heifers were assigned to P₄ classes based on P₄ concentrations in serum samples collected just before treatment with the first GnRH injection (d 0), treatment with PGF_{2 α} (d 6) and treatment with the second GnRH injection (d 8) as described by Cordoba and Fricke (2002) with the following modifications. For samples collected just before the first 2 injections, serum samples were classified as either low (L; ≤ 1.0 ng/mL) or high (H; > 1.0 ng/mL) at the time of each injection. For serum samples collected on d 8, a decrease of 60% or more of the P₄ concentration at treatment with PGF_{2 α} by 48 h after treatment was additionally considered to define low P₄. This second criterion for defining low P₄ on d 8 was included based on a preliminary study in which P₄ was measured after induced luteolysis in heifers using PGF_{2 α} preceded 6 d by GnRH (Rivera and Fricke, 2002).

This classification system resulted in 8 possible P₄ class permutations for GPG heifers: HHH, HHL, HLH, HLL, LHH, LHL, LLH, and LLL. Only (82%, 144/175) of GPG heifers were assigned to P₄ classes because blood sampling was discontinued for the 31 heifers receiving AI after removed tail chalk before scheduled TAI (Table 1). The P₄ classes were used to estimate the stage of the cycle at initiation of GPG, response to the first GnRH injection, and luteolysis in response to

Table 1. Reproductive parameters for GPG heifers receiving timed AI based on progesterone (P₄) concentrations of blood samples collected immediately before each injection of the synchronization protocol.

P ₄ Class ¹	n	CL regression ² n (%)	Ovulation after 2nd GnRH ³ n (%)	Synchronization rate ⁴ n (%)	PR/AI ⁵ n (%)	Pregnancy loss ⁶ n (%)
High P ₄ on d 8						
HHH	3	0 (0.0)	2 (66.7)	0 (0.0)	1 (33.3)	0 (0.0)
LHH	1	0 (0.0)	1 (100.0)	0 (0.0)	0 (0.0)	0 (0.0)
HLH	0	—	—	—	—	—
LLH	0	—	—	—	—	—
Total	4	0 (0.0)	3 (75.0)	0 (0.0)	1 (25.0)	0 (0.0)
Low P ₄ on d 8						
LLL	0	—	—	—	—	—
HLL	12	—	11 (91.6)	0 (0.0)	4 (33.3)	0 (0.0)
LHL	34	34 (100.0)	32 (94.1)	32 (94.1)	11 (32.3)	2 (18.2)
HHL	94	94 (100.0)	85 (90.4)	85 (90.4)	39 (41.5)	3 (7.7)
Total	140	128 (91.4)	128 (91.4)	117 (83.6)	54 (38.6)	5 (9.3)
Overall	144	128 (88.9)	131 (91.0)	117 (81.2)	55 (38.2)	5 (9.0)

¹Combinations of highs (H; > 1 ng/mL) and lows (L; ≤ 1 ng/mL) represent serum progesterone (P₄) concentration at the first GnRH injection (d 0), the PGF_{2α} injection (d 6), and the second GnRH injection (d 8) of the GPG protocol.

²CL regression was considered to occur when serum P₄ concentration decreased to ≤ 1 ng/mL or there was a decrease of 60% or more of the P₄ concentration at treatment with PGF_{2α} by 48 h after treatment.

³Number of heifers that ovulated at least one follicle by 48 h after the second GnRH injection expressed as a percentage of heifers receiving the GPG protocol. Double ovulation rate was 5% (7/131).

⁴Number of heifers with low P₄ on d 8 and in which a follicle ovulated within 48 h of the second GnRH injection expressed as a percentage of heifers receiving the GPG protocol.

⁵Proportion of heifers diagnosed pregnant at 30 d after TAI.

⁶Proportion of heifers diagnosed pregnant 30 d after TAI that were diagnosed nonpregnant 75 d after TAI.

PGF_{2α} for heifers receiving the GPG protocol similar to the methods described by Moreira et al. (2001) and Cordoba and Fricke (2002).

Ultrasonography

To assess ovulatory response to the second GnRH injection of the GPG protocol, ovarian structures were monitored at the time of the second GnRH injection and 48 h thereafter for heifers in the GPG group receiving TAI by using an ultrasound machine equipped with a transrectal 7.5 MHz linear-array transducer (Aloka 500V; Corometrics Medical Systems, Inc., Wallingford, CT). Ultrasound examinations were not conducted for GPG heifers that received AI after removed tail chalk before scheduled TAI or for TC heifers. During each ultrasound examination, a sketch of the location and diameter of all ovarian structures ≥ 8 mm in diameter was recorded. For each ovarian structure, 2 diameter measurements taken at right angles were recorded on a single frozen image of the apparent maximal diameter of the structure using the digital calipers of the ultrasound machine, and diameters were calculated as the mean of these 2 measurements. Ovulatory response after the second GnRH injection was determined by the presence of one or more dominant follicles at the time

of the second GnRH injection and the absence of one (single ovulation) or 2 (double ovulation) of those follicles at an ultrasound examination conducted 48 h later (Fricke et al., 1998).

Heifers with a serum P₄ concentration > 1 ng/mL just before treatment with PGF_{2α} were considered to have a functional CL. Luteal regression was considered to occur when serum P₄ concentration decreased to ≤ 1 ng/mL or there was a decrease of 60% or more of the P₄ concentration at treatment with PGF_{2α} by 48 h after treatment. Double ovulation rate was calculated as the number of heifers that ovulated 2 follicles within 48 h of the second GnRH injection and expressed as percentage of heifers that ovulated after the second GnRH injection. Synchronization rate was defined as the proportion of heifers that underwent luteal regression (P₄ < 1.0 ng/mL or 60% reduction as described previously) by the second GnRH injection and in which ovulation occurred by 48 h after the second GnRH injection expressed as percentage of the total number of heifers receiving the protocol.

Pregnancy Rate per Artificial Insemination

Pregnancy status was determined 30 and 75 d after d 8 of the experiment (i.e., the day that GPG heifers

received TAI) using transrectal ultrasonography. Due to variation in the interval from the start of the experiment to first AI among TC heifers, mean ($\pm$ SEM) interval for all heifers from first insemination to pregnancy diagnosis was 31.5 ± 0.1 and 74.5 ± 0.3 d after first AI. Visualization of a fluid-filled uterine horn and the presence of a conceptus were used as positive indicators of pregnancy (Fricke et al., 1998). Pregnancy rate per artificial insemination (PR/AI) to first AI at 31.5 ± 0.1 and 74.5 ± 0.3 d after AI and pregnancy loss between these 2 stages were calculated for heifers in both treatments. Cumulative pregnancy rate for all AI during the breeding period also was calculated for all heifers in both treatments. Fetal sex was determined for all pregnant heifers at the ultrasound examination conducted on d 74.5 ± 0.3 of gestation based on location of the genital tubercle. A fetus was recorded as a male when the genital tubercle was located in the area caudal to the umbilicus and a female when located beneath the tail (Curran and Ginther, 1991).

Body Size Parameters

Body condition score was assigned to each heifer at first AI by the same individual throughout the experiment using a quarter-point scale from 1 to 5, where 1 = emaciated and 5 = obese (Ferguson et al., 1994). Live BW was estimated for each heifer at first AI using a commercial dairy tape (Weight-By-Breed Dairy Cow Tape; Nasco, Fort Atkinson, WI), and hip height and wither height were measured for each heifer at first AI on a level concrete surface with the heifer in a normal standing position. Wither height index (WHI) was calculated as BW divided by wither height as described by Looper and Bethard (2000), but using metric rather than Imperial units of measure.

Statistical Analyses

Categorical variables (ovulatory response after GnRH and PR/AI) were analyzed by multifactorial logistic regression using the LOGISTIC procedure of SAS (SAS, 1999). The model used to analyze PR/AI to first AI included treatment, inseminator, replicate, and the interaction of treatment $\times$ inseminator, with BCS, wither height, hip height, and BW at first AI as regression variables. Because all heifers within a replicate were turning 13 mo of age within a 2-wk period, age was not included in this model. Continuous variables (days to first AI, days to conception, follicular diameter) were analyzed using the GLM procedure of SAS. Survival analysis (i.e., failure time analysis) was used to assess the effect of treatment on time to first AI and time to pregnancy by using the Log-Rank method of

the LIFETEST procedure of SAS. Observations from heifers that were not inseminated (for analysis of time to first AI) or failed to become pregnant (for analysis of time to pregnancy) during the AI breeding period were considered as censored. For all analyses, a significant difference was considered to occur when $P \leq 0.05$, whereas a statistical tendency was considered to occur when $P > 0.05$ and ≤ 0.15 .

RESULTS AND DISCUSSION

Accuracy of Detection of Estrus

To evaluate the accuracy of the tail chalk system for detection of estrus and to determine the proportion of GPG heifers not in estrus when receiving AI after removed tail chalk before scheduled TAI, blood samples were collected from heifers on the day of AI regardless of treatment during the first 8 d of the experiment. It was assumed that heifers with low P_4 were likely in estrus; however, it is possible that some heifers had low P_4 and were not in estrus (i.e., anovulatory animals). Nevertheless, our objective was to determine the proportion of heifers that were not in estrus at the time of insemination (i.e., high P_4 at AI). Only 8.2% of heifers receiving AI had a serum P_4 concentration ≥ 1 ng/mL on the day of AI, whereas 91.8% of heifers had a serum P_4 concentration < 1 ng/mL on the day of AI (Table 2). No heifers with high P_4 on the day of AI were pregnant ~ 30 d after that service, whereas heifers with low P_4 on the day of AI had a PR/AI at ~ 30 d after AI similar to that of the 172 heifers receiving AI after removed tail chalk (Tables 2 and 3).

Although the correct timing of AI cannot definitively be established for heifers with low P_4 on the day of AI, these heifers were inseminated in the absence of a functional CL, a stage of the cycle when estrus and ovulation would normally occur. By contrast, heifers with high P_4 on the day of AI had a functional CL at AI and were inseminated at an incorrect stage of the cycle. Thus, use of tail chalk in the present study was an effective method for estrus detection for first AI resulting in an overall accuracy of 91.8%. Cordoba and Fricke (2002) reported an 84% submission rate to first AI during a 23-d AI breeding period using a tail paint system in lactating dairy cows managed in a grazing-based system, and Macmillan et al. (1988) reported a 95% submission rate using a similar tail painting system in heifers after synchronized estrus.

Effect of Treatment on PR/AI

Overall PR/AI to first AI for all heifers in the present study was 42.4% (147/347) and, although PR/AI did not differ statistically between treatments, PR/AI tended

Table 2. Accuracy of timing of artificial insemination and pregnancy rate per artificial insemination (PR/AI) during the first 8 d of the AI breeding period for Holstein dairy heifers in both treatments receiving AI after removed tail chalk.

Day	Heifers inseminated	Progesterone status ¹ at AI		Accuracy ² %
		High n (PR/AI, %) ¹	Low n (PR/AI, %)	
1	18	1 (0.0)	17 (29.4)	94.4
2	7	0 (0.0)	7 (57.1)	100.0
3	9	2 (0.0)	7 (29.4)	77.8
4	3	0 (0.0)	3 (0.0)	100.0
5	15	2 (0.0)	13 (53.8)	86.7
6	14	0 (0.0)	14 (42.8)	100.0
7	20	2 (0.0)	18 (61.1)	90.0
8	11	1 (0.0)	10 (40.0)	90.9
Overall	97	8 (0.0)	89 (44.9)	91.8

¹Serum progesterone concentration of blood samples collected on the day of AI was classified as either high (> 1 ng/mL) or low (≤ 1 ng/mL).

²Proportion of heifers receiving AI that had low serum P₄ concentration on the day of AI.

to be greater ($P = 0.12$) for TC than for GPG heifers (Table 3). One factor explaining the tendency for a lower PR/AI in GPG heifers may be due to the proportion of GPG heifers that ovulated a follicle after the second GnRH injection. Studies in which ovulation rate after the second GnRH injection of Ovsynch was assessed in lactating dairy cows using a similar technique were 87% (Vasconcelos et al., 1999) and 84% (Fricke et al., 1998). In the present study, ovulation rate after the second GnRH injection for heifers receiving TAI was 91.0% (Table 1). It is important to note that this figure

does not include the 31 heifers that received AI based on removed tail chalk before TAI. Ovulation rate after the second GnRH injection would have decreased dramatically had these heifers not been detected in estrus during the protocol, and this is likely the reason that heifers respond poorly to Ovsynch when managed as a TAI protocol without detection of estrus (Pursley et al., 1997). Adjusting the PR/AI of 38.2% (55/144) for all GPG heifers in the present study to include only the 131 heifers that ovulated after the second GnRH injection would increase PR/AI to 42.0% (55/131, Table 1), a PR/

Table 3. Effect of treatment on pregnancy rate per artificial insemination (PR/AI), pregnancy loss, and cumulative pregnancy rate of Holstein dairy heifers during the 42-d AI breeding period.

Item	Treatment		P-value
	TC	GPG	
PR/AI to 1st AI at 30 d after TAI ¹			
Overall, %	46.5	38.3	0.12
(no./no.)	(80/172)	(67/175)	
After TAI, %	—	38.2	—
(no./no.)		(55/144)	
After removed tail chalk, %	46.5	38.7	0.44
(no./no.)	(80/172)	(12/31)	
PR/AI to first AI at 75 d ¹			
Overall, %	41.9	34.3	0.15
(no./no.)	(72/172)	(60/175)	
Pregnancy loss, 30 to 75 d ¹			
Overall, %	10.0	10.4	0.98
(no./no.)	(8/80)	(7/67)	
PR/AI to 2nd and 3rd AI at 41 to 66 d after AI			
Overall, %	34.0	42.1	0.25
(no./no.)	(31/91)	(51/121)	
Cumulative pregnancy rate ²			
Overall, %	58.2	63.4	0.38
(no./no.)	(103/177)	(111/175)	

¹Due to variation in the interval from the start of the experiment to first AI among TC heifers, mean (± SEM) interval for all heifers from first insemination to pregnancy diagnosis was 31.5 ± 0.1 and 74.5 ± 0.3 d after first AI.

²Proportion of heifers diagnosed pregnant to AI during the 42-d AI breeding period.

AI similar to that observed for TC heifers (46.5%, Table 3).

Pregnancy loss contributes to reproductive inefficiency in cattle because fertility assessed at any point during pregnancy is a function of both conception rate and pregnancy loss (Fricke, 2002). Fertilization rate after AI in beef cows is 90%, whereas embryonic survival is 93% by d 8 and only 56% by d 12 after AI (Diskin and Sreenan, 1980). Reported pregnancy losses in lactating dairy cows are greater than that in dairy heifers (20 vs. 5%; Smith and Stevenson, 1995). The 10.2% pregnancy loss in the present study is greater than that reported by Smith and Stevenson (1995) but less than the 20.5% (Smith and Stevenson, 1995) and the 13.5% (Fricke et al., 1998) reported during a comparable stage of pregnancy in lactating dairy cows.

Follicles in lactating dairy cows achieve ovulatory capacity around 10 mm in diameter (Sartori et al., 2001). For GPG heifers in the present study, diameter of the largest follicle at the time of the second GnRH injection affected ovulatory response. Follicle diameter at GnRH was greater ($P < 0.01$) for follicles that went on to ovulate by 48 h after the second GnRH injection than for follicles failing to ovulate (13.2 ± 0.2 vs. 10.2 ± 0.8 mm, respectively). By contrast, follicle size at the time of the second GnRH injection did not differ ($P = 0.37$) among GPG heifers based on pregnancy status assessed 30 d after TAI.

Effect of Inseminator on PR/AI

The overall PR/AI of 42.4% for the 347 heifers receiving AI in the present study was lower than expected. Although fertility of dairy heifers is consistently higher than that of lactating dairy cows (Ron et al., 1984), the reported PR/AI for dairy heifers varies widely among studies. Fertility of heifers receiving AI at standing estrus was reported to be 47 (Donovan et al., 2003), 59 (Gwazdauskas et al., 1981), 55 to 66 (Butler and Smith, 1989), and 74% (Pursley et al., 1997). One reason for the poor overall PR/AI in the present study was the poor performance of 2 of the 3 herd inseminators that conducted AI throughout the study (Table 4). Most herd-level variation in conception rate among heifers is due to variation among inseminators (Ron et al., 1984). Barth (1993) reported that timing of AI, semen quality, semen handling, and inseminator expertise influenced fertility to AI in cattle. Improper semen placement in the reproductive tract also affects fertility to AI with over half of inseminators evaluated depositing semen into the cervix rather than correctly placing it into the uterine body (López-Gatius, 1999). A common mistake was for inseminators to deposit semen in the

cervix while withdrawing the pipette or straw during AI (Zavy and Geisert, 1994).

When PR/AI for the 3 inseminators in the present study was compared, the 2 herd employees had PR/AI of 24.8 and 30.0% (inseminators 1 and 2, respectively), which were lower ($P < 0.01$) than that of the herd manager (inseminator 3) who had a PR/AI of 58.0% (Table 4). No treatment $\times$ inseminator interaction was detected ($P = 0.70$), indicating that treatment effects on PR/AI in the present study were not confounded by the inseminator effect. A similar effect of AI technician on conception rate has been reported for beef heifers synchronized using melengestrol acetate and PGF_{2 α} (Salverson et al., 2001). Had the herd manager inseminated all heifers in the present experiment, PR/AI would have been acceptable based on previous reports in dairy heifers (Gwazdauskas et al., 1981; Butler and Smith, 1989).

Reproductive Performance During the AI Breeding Period

Research has consistently supported that lifetime milk yield, 305-d lactation yield, and lifetime profit of replacement heifers are maximized when heifers calve for the first time between 23 and 25 mo of age (Heinrichs, 1993; Van Amburgh et al., 1998) and weigh around 560 kg after calving (Hoffman, 1997; Loooper and Bethard, 2000). Age at first breeding coupled with reproductive efficiency to first and subsequent breedings determine average age at first calving and, more important, the variation in age at first calving among similar-aged heifers in a breeding group (Fricke, 2003). In the present study, GPG heifers were synchronized and received first AI by d 8 of the AI breeding period. This strategy not only increases the AI service rate and groups heifers to conceive more uniformly at the onset of the breeding period, but also concentrates expression of return estrus for heifers failing to conceive to first AI (see Figure 2).

As expected, 17.7% (31/175) of GPG heifers received AI after removed tail chalk before scheduled TAI (Table 3; Figures 1 and 2). Two of these heifers had high P_4 on the day of AI; therefore, the proportion of heifers correctly identified in estrus before TAI was 17.0% (29/175), and these heifers received AI on $d 5.3 \pm 0.2$ of the AI breeding period. In 2 previous reports, 17% of randomly cycling dairy (Rivera and Fricke, 2002) and beef (Roy and Twagiramungu, 1996) heifers expressed estrus during a similar synchronization protocol as that tested in the present study. These heifers likely received the first GnRH injection during late diestrus or early proestrus (i.e., d 14 to 18) and failed to synchronize in a similar physiologic scenario as described in detail by Cordoba and Fricke (2002) for lactating cows receiv-

Table 4. Effect of inseminator on pregnancy rate per artificial insemination (PR/AI) of Holstein dairy heifers receiving AI after removed tail chalk (TC) or synchronization of ovulation and timed artificial insemination (GPG).

Treatment	Inseminator		
	1	2	3
TC, %	30.2	33.3	62.6
(no./no.)	(16/53)	(12/36)	(52/83)
GPG, %	20.0	25.0	53.8
(no./no.)	(12/60)	(6/24)	(49/91)
Overall, %	24.8 ^a	30.0 ^a	58.0 ^b
(no./no.)	(28/113)	(18/60)	(101/174)

^{a,b}Within a row, percentages with different superscripts differ ($P < 0.01$). Treatment by inseminator interaction was not significant ($P = 0.70$).

ing Ovsynch. The proportions of dairy heifers with 2, 3, and 4 follicular waves were 56, 33, and 11%, respectively (Sartori et al., 2004), indicating that nearly half of dairy heifers have more than 2 follicular waves per estrous cycle. In a previous study, all dairy heifers ini-

tiating Ovsynch on d 15 of the estrous cycle were observed in estrus before the second GnRH injection, whereas estrus behavior was not observed when the protocol was initiated on d 2, 5, 10, or 18 of the cycle (Moreira et al., 2000a). For heifers exhibiting 3 follicular waves, the later stage of the cycle coincides with emergence of the third follicular wave (Ginther et al., 1996) when the dominant follicle of the second wave has lost ovulatory capacity and a follicle from the third wave has not yet undergone deviation and acquired ovulatory capacity (Sartori et al., 2001). By contrast, heifers exhibiting 2 follicular waves exhibit a dominant follicle with ovulatory capacity at this time (Ginther et al., 1996) and would not exhibit premature estrus and ovulation due to the presence of P_4 from a GnRH-induced CL.

Interval to first AI was greater ($P < 0.01$) for TC than for GPG heifers with 45.8% (81/177) of TC heifers and 100.0% (175/175) of GPG heifers receiving first AI by d 8 of the AI breeding period (Figure 1, upper panel). All 175 GPG heifers received at least one AI service, whereas 2.8% (5/177) of TC heifers did not receive an AI service during the 42-d AI breeding period (Figure 1, upper panel). Despite the reduced time to first AI, survival time to pregnancy during the 42-d AI breeding period (Figure 1, lower panel) did not differ ($P = 0.65$) between treatments. By the end of the 42-d AI breeding period, 58.2% (103/177) of TC heifers had become pregnant compared with 63.4% (111/175) of GPG heifers (Table 3).

The frequency distributions for the number of heifers receiving second and third AI, during the 42-d AI breeding period are shown in Figure 2. Mean days to first AI was greater ($P < 0.01$) for TC than for GPG heifers (9.9 ± 0.6 vs. 7.5 ± 0.01 d). In the present study, the GPG protocol was initiated on the first day of the AI breeding period and all heifers in this group were inseminated by d 8. Under these management conditions, heifers not conceiving to TAI are likely to display estrus and receive a second AI by d 19 to 24 after TAI. Because

Figure 1. Effect of treatment on survival to first AI service (upper panel) and survival to pregnancy (lower panel) during the 42-d AI breeding period. Heifers were randomly assigned to receive either synchronization of ovulation and timed AI and AI after removed tail chalk for the entire AI breeding period (GPG; $n = 175$), or AI after removed tail chalk for the entire AI breeding period (TC; $n = 177$). A treatment effect ($P < 0.01$) was detected for d to first AI service (upper panel), whereas treatment did not affect d to pregnancy (lower panel).

Figure 2. Frequency distribution for heifers receiving second or third AI services. Heifers were randomly assigned to receive either synchronization of ovulation and timed AI and AI after removed tail chalk for the entire AI breeding period (GPG; $n = 175$), or AI after removed tail chalk for the entire AI breeding period (TC; $n = 177$). Day 8 of the AI breeding period = TAI for GPG heifers.

average estrous cycle duration in dairy heifers is 22 d (Sartori et al., 2004), TC heifers on d 0 or 1 of the estrous cycle at the breeding period onset likely would not have the opportunity for a second AI service during the 42-d AI breeding period. Thus, not only is less time needed for estrus detection, but service rate is also increased in GPG heifers for both first and second AI, and the frequency distributions of second and third AI illustrate these trends (Figure 2).

Overall, 8.3% of pregnant heifers (7.3%, 6/82 of TC heifers and 9.1%, 10/109 of GPG heifers) mistakenly received an AI service based on tail chalk removal after first AI but before pregnancy diagnosis at ~30 d after first AI. This was based on the age of the fetus assessed using ultrasonography at pregnancy diagnosis and the time intervals elapsing between the first and second recorded AI services. The 8.3% error rate in AI for repeat services agrees with the 91.8% accuracy we reported for heifers submitted for first AI (Table 2). Artificial insemination during pregnancy has been thought to induce pregnancy loss either through mechanical trauma to fetal membranes or the introduction of infection into the uterine environment (Noakes et al., 2001). Macmillan et al. (1977) reported that an unnecessary reinsemination during diestrus reduced the PR to a previous insemination at the correct time by over 30%.

In the present study, mistaken AI services cannot fully account for pregnancy loss because only 20% (3/15) of the heifers experiencing loss received AI after first AI but before pregnancy diagnosis.

GPG Heifers with High Progesterone on d 8

Heifers with high P_4 on d 8 failed to synchronize luteal function in response to the GPG protocol. Overall, the proportion of heifers with high P_4 on d 8 was only 2.7% (4/144), and 75% of these heifers ovulated after the second GnRH injection (Table 1). The observation that one of the 4 heifers with high P_4 on d 8 (2.7 ng/mL) was diagnosed pregnant at 30 d after TAI illustrates the limitation of this P_4 classification scheme for defining reproductive function.

Heifers in the HHH and LHH classes likely had a functional CL at the time of $\text{PGF}_{2\alpha}$ treatment that failed to regress in response to $\text{PGF}_{2\alpha}$ treatment. Incomplete luteal regression, defined as serum P_4 concentration > 2.0 ng/mL 48 h after $\text{PGF}_{2\alpha}$, occurred in 7% of lactating dairy cows receiving Ovsynch (Moreira et al., 2001). Rivera and Fricke (2002) reported 20% incomplete luteal regression in dairy heifers when $\text{PGF}_{2\alpha}$ was given 6 d after GnRH, as evaluated by blood samples collected every 24 h after treatment with $\text{PGF}_{2\alpha}$. In dairy heifers,

CL are responsive to $\text{PGF}_{2\alpha}$ at an earlier stage of the estrous cycle compared with lactating cows (Momont and Seguin, 1984). This observation supports our strategy to decrease the interval between GnRH and $\text{PGF}_{2\alpha}$ from 7 to 6 d to adjust for the more rapid turnover of follicular waves in dairy heifers compared with lactating cows (Pursley et al., 1997). No heifers with high P_4 on d 8 displayed either the HLH or LLH profiles. These P_4 classes likely represent heifers that initiated the first GnRH injection during late diestrus or proestrus of the estrous cycle, which would have received AI after removed tail chalk during the GPG protocol in the present study.

GPG Heifers with Low Progesterone on d 8

Heifers with low P_4 on d 8 lacked a functional CL at the second GnRH injection, and these P_4 classes included heifers that were successfully synchronized by the GPG protocol. Of the 144 heifers assigned to a P_4 class, 97.2% (140/144) had low P_4 on d 8 (Table 1). The rate of luteal regression in these heifers was 91.4% (128/140), ovulation rate to the second GnRH injection was 91.4% (128/140), and the double ovulation rate for the heifers that ovulated after GnRH was 5.5% (7/128). Overall, 83.6% (117/140) of these heifers synchronized follicular and luteal function, resulting in a PR/AI of 38.6% (Table 1).

No heifers were classified into the LLL profile, indicating that all heifers ovulated before or during the protocol. The 12 heifers in the HLL class were likely in late diestrus or early proestrus at initiation of the GPG protocol similar to HLH and LLH heifers, and some of these heifers may have been in estrus during the protocol but were not detected with removed tail chalk. Furthermore, 33.3% (4/12) of these heifers were diagnosed pregnant 30 d after TAI. One objective of this protocol was to induce heifers to have a functional CL at $\text{PGF}_{2\alpha}$ treatment and a follicle with ovulatory capacity 2 d later at the second GnRH injection. The number of heifers with the desired P_4 profile (HHL) was 65.3% (94/144), expressed as a proportion of the animals receiving TAI (Table 1).

Results from the present study suggest that presynchronization of dairy heifers so that initiation of the GPG protocol would coincide with early to mid diestrus may be beneficial. Presynchronization strategies using 2 injections of $\text{PGF}_{2\alpha}$ have been shown to improve fertility to Ovsynch and TAI in lactating dairy cows (Moreira et al., 2001; Navanukraw et al., 2004). Thus, presynchronization of dairy heifers to reduce or eliminate the HLH, LLH, and HLL profiles may reduce or eliminate the number of heifers in estrus during the protocol, thereby allowing for TAI without estrus detection. In-

creasing the proportion of heifers classified as HHL profile may also maximize fertility to TAI. Further research is needed to improve the response of dairy heifers to the GPG protocol reported in the present study.

Effect of Body Size Parameters

In the present study, heifers were evaluated using both WHI and BCS, to avoid a possible bias due to visual errors in BCS (Looper and Bethard, 2000). Body size parameters and WHI were within body size recommendations by age reported for growing dairy heifers (Hoffman, 1997; Looper and Bethard, 2000) and did not differ for heifers based on pregnancy status ~30 d after first AI supporting the observation that animal-level factors are not strongly associated with fertility in dairy heifers (Donovan et al., 2003).

Fetal Sex Ratio

Fetal sex ratio (proportion of females) assessed at d ~75 of gestation was 50% for TC heifers vs. 54% for GPG heifers. By contrast, the sex ratio in a large population of Holstein cattle in Saudi Arabia was skewed toward male for multiparous cows (46% females). Despite the higher number of females fetuses resulting from TAI, fetal sex ratio for pregnancies resulting from TAI did not differ ($P < 0.43$) from the sex ratio of fetuses from heifers receiving AI after removed tail chalk. Increasing the interval from TAI to ovulation in lactating cows has been reported to increase the number of female calves born (Pursley et al., 1998). In addition, heifers in New Zealand that were synchronized had more female calves (53.8%) compared with untreated control heifers (45.7%; Xu and Burton, 1999). The mechanism by which this skewing of the sex ratio occurs is not known. In the present study, we assessed fetal gender ratio at ~75 d of gestation, whereas some of the effects of previous studies may have occurred due to selective survival of females or loss of males during gestation. Further research is needed to assess the effect of timing of AI using the GPG protocol described in the present study on fetal sex ratio in dairy heifers.

CONCLUSION

The GPG synchronization protocol assessed in the present study can yield acceptable reproductive performance in dairy heifers when estrus detection and AI are performed during the protocol and AI efficiency is optimized. Such a protocol may be useful for dairy producers who wish to breed a group of heifers within an 8-d period and reduce the need for extended periods of estrus detection. Furthermore, use of this protocol will

concentrate return services for heifers failing to conceive to first AI and may allow for development of systematic resynchronization protocols for submitting heifers for second AI. Expression of estrus during the protocol greatly reduces the synchronization response to the GPG protocol. Hence, future studies should focus on increasing the proportion of heifers receiving TAI and reducing the necessity for estrus detection. Such a strategy may involve hormonal presynchronization to place heifers in an optimal stage of the estrous cycle at the onset of the protocol, or the inclusion of progestin treatment during the protocol to suppress expression of estrus before scheduled TAI.

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