The complex of hypericin with β -lactoglobulin has antimicrobial activity with potential applications in dairy industry

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ABSTRACT

Using a combination of molecular modeling and spectroscopic experiments, the naturally occurring, pharmacologically active hypericin compound is shown to form a stable complex with the dimeric form of β -lactoglobulin (β -LG). Binding is predicted to occur at the narrowest cleft found at the interface between monomers in the dimeric β -LG. The complex is able to preserve the fluorescence and singlet oxygen photosensitizing properties of the dye. The equilibrium constant for hypericin binding has been determined as $K_a = 1.40 \pm 0.07 \mu M^{-1}$, equivalent to a dissociation constant, $K_d = 0.71 \pm 0.03 \mu M$. The complex is active against *Staphylococcus aureus* bacteria. Overall, the results are encouraging for pursuing the potential application of the complex between hypericin and β -LG as a nanodevice with bactericidal properties for disinfection.

Key words: hypericin, β -lactoglobulin, photosensitizer, antimicrobial

INTRODUCTION

Antimicrobial products are important for the health and welfare of livestock, but prudent use is necessary. In fact, the overuse of antibiotics can generate drug-resistant bacteria, with the possibility of transferring them to the food chain (IDF, 2013). For example, methicillin-resistant *Staphylococcus aureus* (MRSA) is a well-known food-borne pathogen that produces heat-stable enterotoxins during growth on a variety of foods and is an important regional cause of human infection (Doyle et al., 2011). Recently, it has been noted

that livestock animals can be reservoirs of MRSA and transmit it to humans that live in close contact with them; MRSA from animals is called livestock-associated MRSA (Graveland et al., 2011). For this reason, it is important to develop new alternative strategies for preventing and treating infectious animal diseases of bacterial origin (Trevisi et al., 2014). Indeed, development of novel methods for decontamination of food processing and handling environment is a key topic for food science (Demirci and Ngadi, 2012; Kairyte et al., 2012; Luksiene and Brovko, 2013).

Antibacterial photosensitization-based treatment (APBT) is an emerging methodology that relies on the combined action of an otherwise nontoxic molecule (called photosensitizer), visible light, and oxygen to produce cytotoxic effects by the photoinduced generation of reactive oxygen species (photodynamic effect), particularly singlet oxygen (1O_2). Thus, photoexcitation of the photosensitizers leads to the formation of 1O_2 , a nonradical, electronically excited form of the dioxygen molecule that is highly reactive against a vast array of cellular components ranging from membrane lipids to proteins and nucleic acids (Ogilby, 2010; Jori et al., 2011). The APBT has proved to be valuable for the treatment of localized microbial infections, effectively acting on several classes of microbial pathogens without inducing insurgence of photoresistant species even after multiple treatments. The selectivity of microbial cell killing in comparison with the constituent host tissues, and the possibility to activate photodynamic sensitizers by means of inexpensive and safe visible light sources, are further elements in favor of the methodology (Jori et al., 2011).

Hypericin (Hyp; Figure 1) is a natural product, structurally belonging to the chemical class of naphthodianthrone, which is found in plants from the genus *Hypericum* (e.g., St. John's wort; Brockmann et al.,

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1939; Durán and Song, 1986). St. John's wort infusions are being used as herbal aids for depression and Hyp has also been used as an antiviral (Jacobson et al., 2001; Kubin et al., 2005), antibacterial (Kairyte et al., 2012; Yow et al., 2012), and antifungal agent (Rezusta et al., 2012). The antimicrobial photoactivity of Hyp against MRSA, both in vitro and in vivo, has been recently assessed (Nafee et al., 2013). Whereas Hyp is largely insoluble in water, it dissolves readily in ethanol or dimethylsulfoxide, where it displays a bright red color due to its absorption and fluorescence emission properties (Durán and Song, 1986; López-Chicón et al., 2012). In these solvents, Hyp is also a strong photosensitizer, with singlet oxygen quantum yields exceeding 0.7 (Michaelli et al., 1993; Fehr et al., 1995). When dissolved in water, Hyp tends to form aggregates, which results in broadening of the bands in the absorption spectrum in a much weaker, broad, and structureless fluorescence emission, and loss of singlet oxygen photosensitizing efficiency (Yamazaki et al., 1993).

Because the effectiveness of Hyp-mediated photodynamic action is heavily dependent on the specific localizing capability of the delivery methodology, proteins appear as particularly interesting delivery agents thanks to their high biocompatibility, bioavailability, and the possibility to select specific proteins suitable to the environment to be targeted. We have recently shown that Hyp binds to the hydrophobic pocket of apomyoglobin, preserving its fluorescence and $^1\text{O}_2$ photosensitizing properties (Comas-Barceló et al., 2013). Furthermore, we have shown that the complex is active against *Staphylococcus aureus* bacteria.

In the current study, we propose the use of β -LG, because it is the most abundant protein in the whey of cow milk (Flower et al., 2000), as a delivery vehicle for antimicrobial photodynamic application of Hyp. β -Lactoglobulin is a small, homodimeric protein of 162 amino acids (~18,400 Da) belonging to the lipocalin

family (Åkerström et al., 2006; Mercadante et al., 2012). Owing to their ability to bind small hydrophobic molecules, while simultaneously being recognized by cell surface receptors, lipocalins were suggested as potential systems for drug delivery (Åkerström et al., 2006). Whereas some lipocalins possess high ligand specificity, others, such as β -LG, demonstrate a remarkable versatility in their ligand-binding patterns. Affinity for small molecules is generally intermediate, with dissociation constants (K_d) around $1\ \mu\text{M}$. We used the B variant of β -LG, characterized by glycine and alanine at positions 64 and 118 (Tanford and Nozaki, 1959), to examine the ability to bind Hyp and the effect of binding on the photodynamic properties of Hyp. Additionally, we show that the complex between β -LG and Hyp is able to inactivate *Staph. aureus* bacteria. In view of the biocompatibility of the β -LG carrier with dairy products as well as with milk processing and handling material, this construct has potential for photodynamic antibacterial applications in the dairy industry.

MATERIALS AND METHODS

Hypericin was obtained from HWI Analytik GmbH (Ruelzheim, Germany). β -Lactoglobulin (isoform B) from bovine milk was obtained from Sigma Aldrich (St. Louis, MO). All samples were used as received.

Docking Calculations

The protein model for the bovine β -LG dimer was derived from the X-ray crystallographic structure (PDB entry 1BEB, <http://www.rcsb.org/>; Brownlow et al., 1997). The structure was refined by including addition of hydrogen atoms using the xLEAP facility in AMBER12 (Case et al., 2012) and the parm99SB (Hornak et al., 2006) force field, removal of nonstandard residues (sulfate anion), and generation of the disulfide bonds Cys66-Cys160 and Cys106-Cys119. Because the N- and C-terminal segments are not observed in the X-ray structure, N-methylamine and acetyl capping groups were added to the C- and N-terminus, respectively. The binding of Hyp was explored by means of docking calculations carried out with GLIDE (Friesner et al., 2004; Halgren et al., 2004). Because the size of Hyp precludes its binding to the cavity shaped in the interior of the lipocalin-type β -barrel, docking was performed in the 2 pockets formed at the interface of the monomers in the dimeric complex. The first pocket included the solvent-exposed residues inside a cube of $39,300\ \text{\AA}^3$, which was centered between side chains of residues Arg148 of one monomer and Asp137 of the other. The second cavity was defined by the exposed residues inside a cube of the same size centered at the midpoint of the segment

that connects the side chains of residue Trp61 in each monomer.

General Spectroscopic Instrumentation

Absorption spectra recorded using a Varian Cary 6000i (Varian Inc., Palo Alto, CA). Fluorescence spectra were recorded using a Spex-Fluoromax 4 (Horiba Jobin Yvon, Edison, NJ) or a Perkin Elmer LS50 spectrofluorometers (PerkinElmer, Waltham, MA).

Singlet Oxygen Detection

Production of the reactive species singlet oxygen was assessed by monitoring its specific phosphorescence at 1,275 nm with a setup described in detail elsewhere (Jiménez-Banzo et al., 2008). Hypericin was excited by a pulsed laser at 532 nm and the emission of singlet oxygen was detected by a dedicated near infrared photomultiplier.

Microbial Strains and Growth Conditions

Staphylococcus aureus CECT 239 was obtained from the Spanish Type Culture Collection (CECT). Bacterial cells were grown overnight in sterile tryptic soy broth at 37°C. Stock inoculum suspensions were prepared in PBS and adjusted to optical densities of 0.4 at 600 nm.

Photodynamic Inactivation of *Staph. aureus*

Cell suspensions in PBS were incubated with the photosensitizer for 30 min in the dark at room temperature. Then, 0.3 mL of the suspensions were placed in 96-well plates. The plates were illuminated from the top by use of light-emitting diode green light for 15 or 30 min (18 and 37 J/cm², respectively), then serially diluted, seeded on tryptic soy agar and incubated for 24 h at 37°C. Experiments were carried out in duplicate for each condition, including cell controls and dark controls. Colony-forming units were counted to calculate the survival fraction.

RESULTS AND DISCUSSION

Modeling of β -LG-Hyp interaction

β -Lactoglobulin forms dimers (2β -LG) under conditions close to those encountered physiologically, with a dimerization constant of $K_D = 8.6 \mu M$ at pH 7.5 (Mercadante et al., 2012). β -Lactoglobulin is shaped to accommodate small hydrophobic compounds, such as retinol and the aliphatic chains of FA, in the lipocalin-type β -barrel. However, the cavity is too narrow to ac-

commodate Hyp, which suggests that this compound should bind other pockets. In particular, inspection of the dimeric structure of β -LG revealed that Hyp might bind to the clefts formed at the interface between monomers (Figure 2). The analysis of the best-ranked poses supported the binding of Hyp to the narrow cleft, as the predicted score (-7.4 kcal/mol) is ~ 2.5 kcal/mol more favorable than the binding to the wide cleft (score of -4.9 kcal/mol). This finding is not unexpected, given the hydrophobicity of the compound and the narrow separation between the walls of this cleft, where Hyp is largely occluded from the solvent. In contrast, binding to the wide cleft leaves one of the 2 faces of Hyp exposed to the solvent (Figure 2).

Spectroscopic Analysis of Hyp Binding to β -LG

Binding between Hyp and 2β -LG can be monitored through the effects on the absorption spectrum and fluorescence emission. Aqueous solutions of Hyp show broad absorption bands and very weak and structureless fluorescence emission (Figure 3A; Miskovsky et al., 1998). When Hyp is dissolved in the presence of 2β -LG, its absorption spectrum shows sharper bands instead and, most importantly, the fluorescence emission increases substantially and becomes structured (Figure 3B). These facts indicate that Hyp is located in a substantially less polar environment than water, as previously observed with apomyoglobin and human serum albumin (Miskovsky et al., 1998; Das et al., 1999; Comas-Barceló et al., 2013), as a result of binding to the protein. These conditions are a prerequisite for preservation of the singlet oxygen photosensitization via energy transfer from the Hyp triplet state. A more detailed characterization of the photophysical properties of the complex is beyond the scope of the present work and will be reported elsewhere.

As virtually only bound Hyp molecules are fluorescent, the recorded fluorescence emission intensity is proportional to the fraction of bound molecules. Accordingly, the affinity of Hyp for 2β -LG can be evaluated by monitoring the fluorescence emission as a function of Hyp concentration (Figure 3C). A model for the binding equilibrium $\text{Hyp} + 2\beta\text{-LG} \rightleftharpoons 2\beta\text{-LG-Hyp}$ (Comas-Barceló et al., 2013) affords an association constant, $K_a = 1.40 \pm 0.07 \mu M^{-1}$, corresponding to a dissociation constant, $K_d = 0.71 \pm 0.03 \mu M$.

Singlet Oxygen Production

Production of the reactive species singlet oxygen was demonstrated by observation of its specific phosphorescence at 1,275 nm. Figure 4 shows the intensity of the singlet oxygen emission of Hyp and of the Hyp- 2β -LG

Figure 2. Best poses of hypericin bound to the narrow and wide clefts obtained from docking calculations. Hypericin is shown as sticks [dark gray (orange) and light gray (blue) in the forms bound to the narrow and wide clefts, respectively]. The backbone of the dimeric species of β -LG is shown in dark gray (green), and the gray area displays the surface of the dimer. Color version available online.

complex in aqueous solutions. Consistent with previous results, it can be clearly observed that the complex has a far superior ability than free Hyp to produce singlet oxygen.

Photodynamic Inactivation of *Staph. aureus*

Hypericin is widely used in antimicrobial photodynamic therapy due to its bactericidal and fungicidal effect (Rezusta et al., 2012; Yow et al., 2012). To assess the corresponding effects of the Hyp-2 β -LG complex, photodynamic inactivation tests were carried out on *Staph. aureus* bacteria. The results were compared with those obtained for free Hyp under the same experimental conditions (Figure 5). It can be seen that the complex is devoid of any dark toxicity, yet it is capable

Figure 3. Absorption (A) and fluorescence emission (B) spectra of hypericin solutions ($7 \mu M$) in dimethylsulfoxide (solid gray), in PBS (solid black), and in PBS in the presence of $100 \mu M$ β -LG (dashed). (C) Fluorescence emission intensity (estimated from the integrated area) as a function of hypericin concentration at β -LG = $100 \mu M$ (circles). Assuming a dimerization constant, $K_D = 8.6 \mu M^{-1}$, this corresponds to 2 β -LG $\approx 40 \mu M$. The solid line is the result of the fit with a model for a single binding site for hypericin on 2 β -LG (Comas-Barceló et al., 2013) and afford an association constant, $K_a = 1.40 \pm 0.07 \mu M^{-1}$, corresponding to a dissociation constant of $K_d = 0.71 \pm 0.03 \mu M$.

Figure 4. Production of singlet oxygen by hypericin and the complex hypericin-2 β -LG in PBS. Concentrations: hypericin ($8 \mu M$) and β -LG ($40 \mu M$). Color version available online.

Figure 5. Light dose effects on *Staphylococcus aureus* photoinactivation by the complex hypericin-2β-LG for hypericin (8 μM) and β-LG (40 μM). Color version available online.

of eradicating bacteria (5 logs) upon exposure to visible light. Although the bacterial phototoxicity of Hyp-2β-LG is not much different from that of free Hyp, a major advantage of the use of the Hyp-2β-LG complex is that it is administered to the cell suspension using a buffered aqueous solution, which is clearly more compatible with the use in a food processing environment.

CONCLUSIONS

The complex between Hyp and β-LG preserves the singlet oxygen photosensitizing properties of Hyp. The nanostructured material is effective against *Staph. aureus* suspensions and can decrease the bacterial load by 5 orders of magnitude (99.999%) upon exposure to visible light. The combination of the superb antimicrobial properties of natural hypericin with the full compatibility of the protein carrier with dairy industry processes suggests that it has the potential to be introduced as an effective disinfectant for food manufacturing and handling materials. Further, this APBT might find an application in the routine of milking.

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