Diet-induced pseudohypoparathyroidism: A hypocalcemia and milk fever risk factor

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ABSTRACT
Subclinical hypocalcemia may affect half of all multiparous cows, and clinical hypocalcemia or milk fever affects approximately 5% of dairy cows each year. This disorder of calcium homeostasis can be induced by several dietary factors. Recent studies implicate high dietary potassium and high dietary cation-anion difference (DCAD) with increased risk of milk fever. The hypothesis tested in this study was that high-DCAD diets fed to prepartum cows reduce tissue sensitivity to parathyroid hormone (PTH), inducing a pseudohypoparathyroid state that diminishes calcium homeostatic responses. Multiparous Jersey cows were fed low- or high-DCAD diets in late gestation, creating a compensated metabolic alkalosis in the high-DCAD cows and a compensated metabolic acidosis in the low-DCAD cows. They then received synthetic PTH injections at 3-h intervals for 48 h. Parathyroid hormone is expected to cause an increase in plasma calcium by increasing renal production of 1,25-dihydroxyvitamin D and increasing bone calcium resorption. Plasma calcium concentration increased at a significantly lower rate in cows fed the high-DCAD diet. Cows fed the high-DCAD diet also produced significantly less 1,25-dihydroxyvitamin D in response to the PTH injections than cows fed the low-DCAD diet. Serum concentrations of the bone resorption marker carboxyterminal telopeptide of type I collagen were numerically lower in cows fed the high-DCAD diet but this difference was not statistically significant. These data provide direct evidence that high-DCAD diets reduce tissue sensitivity to PTH. The metabolic alkalosis associated with high-DCAD diets likely induces a state of pseudohypoparathyroidism in some dairy cows at the onset of lactation, resulting in hypocalcemia and milk fever.

Key words: milk fever, pseudohypoparathyroid, dietary cation-anion difference

INTRODUCTION
Milk fever is the common name given to a severe, acute hypocalcemic condition occurring in 5 to 6% of all dairy cows within days of calving. Subclinical hypocalcemia, defined as plasma Ca concentration <8 mg/dL, affects about 50% of all multiparous cows and 25% of heifers (Reinhardt et al., 2011). Hypocalcemia occurs because the mammary gland demand for Ca at the onset of milk production draws Ca from the plasma and extracellular fluid Ca pool faster than it can be replaced. As cows develop hypocalcemia, they respond rapidly by increasing secretion of parathyroid hormone (PTH). In cows that do not develop subclinical hypocalcemia or the clinical hypocalcemia of milk fever, PTH rapidly activates Ca homeostatic mechanisms such as renal re-absorption of urinary calcium, osteoclastic bone resorption, and increased renal production of the hormonal form of vitamin D, 1,25-dihydroxyvitamin D [1,25(OH)2D] to enhance intestinal absorption of dietary Ca (Ramberg et al., 1984). When Ca homeostatic mechanisms are functioning properly, only a small decline in blood Ca occurs. More severe hypocalcemia occurs when these mechanisms are not rapidly or fully initiated. At one time, it was thought that milk fever occurred as a result of failure of the parathyroid gland to recognize and respond to the hypocalcemia developing at the onset of lactation and that excessively high dietary Ca in the prepartum ration caused the parathyroid gland dysfunction (Capen and Young, 1967). However, it was later discovered that cows developing milk fever have very high concentrations of PTH in their blood (Mayer et al., 1969). In a landmark study, Martig and Mayer (1973) were able to demonstrate that the response of late-gestation cows to exogenous PTH (an increase in blood Ca) was diminished compared with the response elicited by PTH in cows in lactation, but they were not able to discern the cause. In severe cases of “relapsing” milk fever (cows relapsing and becoming recumbent again some hours after the
typical intravenous calcium treatment), it was observed that the secreted PTH in these cows failed to stimulate production of 1,25(OH)\(_2\)D as quickly as it does in cows with less severe hypocalcemia (cows that require only a single intravenous calcium treatment to effect a recovery), suggesting again that the periparturient cow’s tissues could be temporarily refractory to PTH. In that study, the cows were fed a diet with alfalfa hay as the forage, which would be considered a high-DCAD diet (Goff et al., 1989).

Several meta-analyses have been done concerning risk factors for milk fever. Oetzel (1991), working with a limited number of studies at the time, concluded that dietary S content and DCAD using the equation utilizing milliequivalents of (Na + K) – (Cl + S) most accurately reflected the risk of a diet causing milk fever. That analysis indicated that the milk fever incidence was highest when diet Ca was 1.16% and that dietary Ca well above or well below 1.16% had a preventative effect against milk fever. A few years later, Charbonneau et al. (2006) concluded that the DCAD equation utilizing milliequivalents of (Na + K) – (Cl + 0.6 S) was the best predictor of clinical milk fever risk, reducing the importance of dietary S and determining that an important aspect of DCAD manipulation was an alteration of acid-base status. Lean et al. (2006) concluded that the DCAD equation utilizing milliequivalents of (Na + K) – (Cl + S) accurately reflected the risk of a diet causing milk fever and that the risk of milk fever was greatest with diets that had 1.1 to 1.3% Ca. Their analysis pointed out a pivotal role for inadequate diet Mg as a very strong milk fever risk factor. Their models also demonstrated that excessive diet phosphorus was another milk fever risk factor.

Decreasing the DCAD, defined as the difference in the number of milliequivalents of cations (primarily K and Na) and anions (primarily Cl and S) in the diet, improves Ca homeostasis at the onset of lactation (Ender et al., 1971). Ender et al. (1971), Dishington (1975), and Block (1984) demonstrated that adding acidogenic chloride and sulfate anions to the cows’ diet in the final weeks of gestation could prevent milk fever. Goff and Horst (1997) demonstrated that high dietary K increased the risk of a cow developing milk fever. The high dietary K content induced metabolic alkalosis in the cows. They noted that the risk of developing severe hypocalcemia at the onset of lactation was greatest in those cows in a state of metabolic alkalosis. Similar results were observed when Na was added to the preparum diet. The mechanism by which addition of anions to a diet to counteract cations in the diet of a cow enhances Ca homeostasis is still not understood. Leclerc and Block (1989), Goff et al. (1991), and Philippo et al. (1994) provided indirect evidence that physiological functions stimulated by PTH, such as bone resorption and production of 1,25(OH)\(_2\)D, were enhanced in cows fed diets with added anions. Unfortunately, differences in milk production, sequestration of Ca in the mammary gland before calving, inappetence on the day of calving and feed intake after calving, and other metabolic stressors (negative energy, protein imbalance) occur on the first day of lactation. This causes difficulty when trying to discern how and why tissues might become refractory to PTH stimulation in experiments conducted around the time of calving. In this study, we tested the hypothesis that metabolic alkalosis induced by diet can induce resistance of target tissues to PTH stimulation in a late-gestation cow model. Using this model, we were able to isolate the effect of high-DCAD diet separately from other risk factors, such as excessive dietary phosphorus and inadequate diet Mg.

**MATERIALS AND METHODS**

**Animals**

All procedures used on the cows in this study were approved by the US Department of Agriculture-Agricultural Research Service National Animal Disease Center’s Institutional Animal Care and Use Committee. Jersey cows entering their third or greater lactation and in their last 2 mo of gestation, weighing from 490 to 545 kg, participated in this study. They were fed either a low-DCAD (n = 8) or a high-DCAD (n = 8) diet for 2 wk (Table 1) to induce a state of metabolic acidosis or alkalosis before they were treated with PTH (Figure 1). The actual number of days before calving when the PTH treatment was begun averaged 28 d, with a range from 6 d before calving to 41 d before calving. The trial was performed in 4 separate experimental periods using 4 cows with approximately the same expected calving dates (2 cows on each diet treatment) in each experimental period. The diets were formulated by mixing either reagent-grade 36% hydrochloric acid (HCl) or potassium carbonate (K\(_2\)CO\(_3\)) into a basal corn silage–grass hay ration to achieve DCAD of −181 and +188 mEq/kg of DM for the low- and high-DCAD diets, respectively. Cows were housed individually in box stalls and limit fed 8.0 kg of diet/d (DM basis) to ensure complete intake of added anions or cations during the course of the trial. Orts were less than 0.75 kg of DM/d in all cows and consisted primarily of corn cobs and some larger hay fibers.

The diets were also formulated to avoid other dietary factors identified as factors that increase the risk of hypocalcemia, allowing isolation of the effects of DCAD. The diets were 0.76% Ca, well below the levels the meta-analyses cited above have implicated as most likely to
induce milk fever. The diet supplied about 60 g of Ca/d, of which about 39 g is estimated to be absorbable using the NRC (2001) model. These dry cows are estimated to require just 16 g of absorbed Ca/d. The diets were 0.41% Mg, well in excess of NRC (2001) requirements. Energy, MP, trace minerals, and vitamins supplied by the diet were in excess of NRC (2001) requirements.

Baseline blood and urine samples were collected from each cow the day before PTH treatment. Total urine collections were made from 0800 to 2000 h on the day before PTH treatment and again during the first 12 h of PTH treatment from time 0 to 12 h (0800 to 2000 h). Samples of urine elicited by micturition (20 mL of mid-stream urine) were also collected at intervals during the remaining 36 h of PTH treatment.

**PTH Treatment**

The synthetic, N-terminal 1–34 bovine PTH fragment used in this study was 69% pure and provided 4,142 IU of PTH/mg (Peninsula Labs Inc., San Carlos, CA; lot no. 9802082). The powdered PTH was weighed into a polyethylene tube that had been previously rinsed with 4% BSA (essentially fatty acid-free and globulin-free bovine albumin, Sigma Chemical Co., St. Louis, MO) to reduce nonspecific binding of PTH to the vessel walls. The PTH was then solubilized in slightly acidic (0.001 N HCl) 0.85% saline containing 2% cysteine and 2% BSA, again to prevent PTH from nonspecifically binding to syringe walls. The PTH solvent had previously been sterilized by passage through a 45-μm filter. Two-milliliter aliquots of this solution constituting a single dose of PTH for injection were kept frozen at −20°C until use. Just before injection, a dose was removed from the freezer, allowed to thaw at room temperature, and drawn up into a 10-mL syringe containing 7 mL of 0.85% saline. Injections were made into the gluteal muscles using a 1.5-inch, 20-gauge needle. The first dose of PTH given to each cow contained 5 mg of PTH as a priming dose. Additional injections of 3.3 mg of PTH were given every 3 h up to 48 h. Jugular venous heparinized blood samples were obtained from each cow just before PTH treatment and at 3-h intervals during the period of PTH administration. Plasma was obtained by centrifugation and stored at −20°C until assayed. A second jugular venous blood sample was also obtained anaerobically in heparinized glass syringes before the first PTH injection and kept sealed at 4°C while being transported to the laboratory for determination of blood gas concentrations and pH (ABL 505 Blood Gas Analyzer, Radiometer, Copenhagen, Denmark).

### Table 1. Composition of low- and high-DCAD diets fed to late-gestation cows for 2 wk before and during parathyroid hormone treatment

<table>
<thead>
<tr>
<th>Item</th>
<th>Low DCAD</th>
<th>High DCAD</th>
</tr>
</thead>
<tbody>
<tr>
<td>Ingredient, DM basis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>Corn silage, kg/d</td>
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<td>3.73</td>
</tr>
<tr>
<td>Prairie grass hay, kg/d</td>
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<td>0.47</td>
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<tr>
<td>Distillers grains, kg/d</td>
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<td>0.99</td>
</tr>
<tr>
<td>Expeller soymeal 44% CP, kg/d</td>
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<td>0.49</td>
</tr>
<tr>
<td>Beet pulp, kg/d</td>
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<td>1.94</td>
</tr>
<tr>
<td>Salt-vitamin-mineral mix, kg/d</td>
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<td>0.11</td>
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<tr>
<td>Dicalcium phosphate, g/d</td>
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<td>57</td>
</tr>
<tr>
<td>Calcium sulfate-2H2O, g/d</td>
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<td>65</td>
</tr>
<tr>
<td>Hydrochloric acid (36% HCl), g/d</td>
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<td>0</td>
</tr>
<tr>
<td>Potassium carbonate, g/d</td>
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<td>110</td>
</tr>
<tr>
<td>Analysis</td>
<td></td>
<td></td>
</tr>
<tr>
<td>CP, %</td>
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<td>13.8</td>
</tr>
<tr>
<td>RUP, %</td>
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<td>6.5</td>
</tr>
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<td>Ether extract, %</td>
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<td>NFC, %</td>
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<td>1.64</td>
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<td>Ca, %</td>
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<td>0.76</td>
</tr>
<tr>
<td>P, %</td>
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<td>0.40</td>
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<td>Na, %</td>
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<td>0.19</td>
</tr>
<tr>
<td>K, %</td>
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<tr>
<td>Mg, %</td>
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<td>0.41</td>
</tr>
<tr>
<td>Cl, %</td>
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<td>1.40</td>
</tr>
<tr>
<td>S, %</td>
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</tr>
<tr>
<td>DCAD, mEq/kg</td>
<td>−181</td>
<td>188</td>
</tr>
</tbody>
</table>
Analyses

Plasma and urine Ca and Mg concentrations were determined by atomic absorption spectrophotometry (Perkin-Elmer Corporation, 1965). Urine creatinine concentrations were determined by colorimetric assay (Chasson et al., 1961). Urine pH was determined within 20 min of collection (Corning pH meter, Corning, NY). Plasma 1,25(OH)₂D concentrations were determined by radioimmunoassay using iodinated 1,25(OH)₂D₃ as tracer (Hollis et al., 1996) with a commercial kit (Diasorin, Stillwater, MN). Assays were performed so that samples from a cow on each diet treatment were paired in the same assay. This assay has a limit of detection of 2.4 pg/mL, within-assay coefficient of variation of 10.4%, and between-assay coefficient of variation of 11.4%.

Carboxyterminal telopeptide of type I collagen (CTX) is an end-product of cathepsin-K-mediated type I collagen turnover and is used as an index of osteoclast bone resorption activity. Serum levels of epitopes of the CTX released into the blood during osteoclast bone collagen digestion were determined at 0, 15, 33, and 48 h of PTH treatment. Serum CTX concentrations were measured using a one-step ELISA (Osteometer, Biotech, Copenhagen, Denmark). This test is based on 2 specific monoclonal antibodies directed against AA sequences within the CTX. The CTX assay has been used in human samples to assess bone resorption rates and was previously validated for the bovine (Liesegang et al., 1998). For this study, CTX concentrations for all samples were determined in a single assay. As a check of the assay validity, serial dilutions of serum from a 3-wk-old calf (actively resorbing bone during growth and bone remodeling) were included in the assay and the assay performed in a linear fashion to known standards included with the kit until CTX concentrations were <0.1 ng/mL, confirming the validation by Liesegang et al. (1998).

Statistical Analysis

Data on blood Ca, Mg, 1,25(OH)₂D, and CTX concentrations and on urine Ca:creatinine and Mg:creatinine ratios were subjected to repeated-measures ANOVA (SAS 8.2, SAS Institute Inc., Cary, NC). Cow nested within dietary treatment was the subject, and hour of PTH treatment was the repeated measure. The autoregressive covariance structure was used because it yielded the Akaike information criterion that was closest to zero (Littell et al., 1998). The main effects included in the initial model were diet (low DCAD, high DCAD), experimental period (1–4), and hour of PTH treatment. We were concerned that the use of 4 cows in each experimental period might have an effect in that cows that went through the protocol at one time might react differently from cows that were treated with PTH at a different time of replication of the dietary treatments. The full model suggested that experimental period had no significant effect on the outcome of the study; thus, experimental period was dropped from the final model. The final repeated-measures ANOVA model included diet, hour of PTH treatment, and the interaction between diet and hour of PTH treatment as main effects. When ANOVA indicated a significant effect at \( P < 0.05 \), post hoc pairwise testing of differences between least squares means was performed, using the Tukey adjustment for the number of tests performed.

Data from the first 15 h of PTH treatment were examined for the rate of change in Ca, 1,25(OH)₂D, and CTX by determining the trendline between data points at time 0 and 15 h of PTH treatment. The slope of this line was determined for each individual cow and the means across the 2 treatments were compared by Student’s \( t \)-test. The 15-h time point was chosen because after that time PTH administration was inducing hypercalcemia in the low-DCAD cows, which likely initiated Ca homeostatic responses antagonistic to the action of PTH, such as calcitonin secretion, to return plasma Ca to normal levels. Calcitonin would quickly reduce bone osteoclast activity (Zaidi et al., 2002). Hypercalcemia (directly) and calcitonin can also down-regulate the renal production of 1,25(OH)₂D (Hove et al., 1984; Omdahl et al., 2002). Data on total urine Ca excretion, blood gas concentration, and blood and urine pH as affected by diet before the first PTH injection were compared across the 2 treatments by Student’s \( t \)-test.

RESULTS AND DISCUSSION

Diet Effects on Acid-Base Status

After 2 wk on the experimental diets and just before treatment with PTH was initiated, the urine pH and blood pH of cows on the low-DCAD diet were significantly lower (\( P < 0.01 \)) than in cows fed the high-DCAD diet (Figure 2). Blood bicarbonate concentrations were also significantly lower (\( P < 0.01 \)) in the low-DCAD cows (data not shown). The relatively high chloride, low K content of the low-DCAD diet induced a metabolic acidosis, with respiratory compensation driving blood bicarbonate slightly below normal. The relatively low chloride, high K content of the high-DCAD diet
induced a compensated metabolic alkalosis in the cows. Urinary excretion of Ca determined in the 12 h from 0800 to 2000 h the day before PTH treatment was almost 10-fold greater in cows fed the low-DCAD diet (3.59 ± 0.16 g) than in the high-DCAD cows (0.39 ± 0.19 g; \( P < 0.001 \)). Plasma Ca concentration before the initiation of PTH treatment was slightly higher in high-DCAD cows than in low-DCAD cows (\( P < 0.07 \)), which may have been a result of the extra Ca lost to urine in the low-DCAD cows (Figure 3). Higher blood Ca in the high-DCAD cows more likely occurred because alkalosis reduces the fraction of total Ca that is in the ionized state and Ca homeostasis is driven by plasma ionized Ca concentration. It is likely that ionized Ca concentration in both groups of cows was similar and that this could be achieved with lower total Ca in low-DCAD cows (Oberleithner et al., 1982).

Plasma Mg concentrations were above 2.2 mg/dL in both groups, well above the renal threshold level of 1.9 mg/dL (Goff, 1999), indicating that Mg status was adequate in both groups (Figure 4). Urinary Mg loss during the 12-h period before PTH treatment was similar across both diets, again suggesting that Mg status was similar in both groups and not a confounding factor in this study. This is important to point out because a low plasma Mg concentration is known to increase the risk of hypocalcemia in cows. Hypomagnesemia can reduce endogenous PTH secretion (Littledike and Goff, 1987; Rude and Gruber, 2004) and strong evidence exists that it also reduces bone and kidney responsiveness to PTH (Contreras et al., 1982; van de Braak et al., 1987; Rude and Gruber, 2004).

Repeated-measures ANOVA demonstrated significant time, diet, and time × diet interactions on both plasma Ca and 1,25(OH)\(_2\)D concentrations. Cows fed the low-DCAD diet exhibited a significant increase (\( P < 0.05 \)) in plasma Ca concentration from baseline (time 0) concentrations by 6 h of PTH treatment, and plasma Ca continued to increase, peaking at 24 h of PTH treatment and remaining elevated thereafter (Figure 3). The mean slope of the change in blood Ca concentrations during the first 15 h of PTH treatment, before development of marked hypercalcemia, was 0.052 ± 0.011 mg/dL per hour in the low-DCAD diet cows. This was 2.7 times (\( P < 0.05 \)) the rate of plasma Ca change in the high-DCAD diet cows, where Ca increased by 0.019 ± 0.008 mg/dL per hour during the first 15 h of PTH treatment.

Plasma 1,25(OH)\(_2\)D concentration was significantly (\( P < 0.05 \)) increased above baseline (time 0) in low-DCAD cows at 6 h of PTH treatment and continued to increase, peaking at 24 h of PTH treatment and remaining elevated thereafter. In high-DCAD cows, we observed a small but significant spike in plasma 1,25(OH)\(_2\)D concentration at 6 h of PTH treatment,
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but values at 9 and 12 h of PTH treatment were not significantly elevated from baseline. In the high-DCAD cows, the plasma 1,25(OH)2D concentration increased significantly above baseline (time 0) at 15, 24, 30, and 33 h of PTH treatment but were statistically similar to baseline levels at 18, 21, and 27 h of PTH treatment (Figure 5). Plasma 1,25(OH)2D concentrations increased 3.15-fold faster during the first 15 h of PTH treatment in low-DCAD diet cows (4.29 ± 0.64 pg/mL per hour) than in the high-DCAD diet cows (1.36 ± 0.73 pg/mL per hour; P < 0.02). In periparturient cows, feeding high-DCAD diets is associated with decreased renal production of 1,25(OH)2D as hypocalcemia developed (Gaynor et al., 1989; Goff et al., 1991; Phillippo et al., 1994). Failure to produce adequate 1,25(OH)2D would be expected to reduce the ability to utilize diet Ca efficiently. In one of the most comprehensive studies on DCAD effects in ruminants conducted, Fredeen et al. (1988) were able to feed high- and low-DCAD diets to goats and were able to conduct Ca absorption studies utilizing Ca45. They determined that low-DCAD diets increased intestinal absorption of Ca (Fredeen et al., 1988).

Only time had a significant effect on osteoclastic bone resorption, as indicated by serum CTX concentration. We observed a significant increase in serum CTX concentration by the 15-h time point in both groups. Only a small additional increase in serum CTX was observed as PTH treatment continued beyond that time in either dietary group (Figure 6). Serum CTX tended to be higher in cows fed the low-DCAD diet but the difference was not statistically significant. The slope of the line drawn based on plasma CTX concentration at 0 and 15 h of treatment indicated that plasma CTX increased in both dietary treatment groups at essentially the same rate (0.020 ± 0.006 vs. 0.015 ± 0.006 ng/mL per hour in the low- and high-DCAD cows, respectively). If a difference in bone resorption activity occurred across these 2 diets, our model lacked the statistical power to discern the difference. However, Fredeen et al. (1988) performed Ca45 tracer studies in goats and found that feeding a low-DCAD diet enhanced release of Ca from bone when the goats were subjected to a calcium challenge. In a periparturient cow model, Leclerc and Block (1989) demonstrated higher serum hydroxyproline concentration in cows fed a low-DCAD diet, and this was associated with improved Ca status at calving.

During the first 12 h of PTH treatment, urinary Ca loss was reduced from 3.59 ± 0.16 to 3.01 ± 0.24 g/12 h in the low-DCAD cows. The low-DCAD cows excreted 0.58 g less Ca during PTH treatment than they did the day before PTH treatment. This represents a 16% reduction in urine Ca with PTH stimulation. In the high-DCAD cows, urine Ca excretion was reduced from 0.39 ± 0.19 to 0.27 ± 0.13 g/12 h during the first 12 h of PTH treatment. The high-DCAD cows excreted 0.12 g less Ca than they did the day before PTH treatment. This represents a 30% reduction in urine Ca with PTH stimulation. As baseline (time 0) urinary Ca excretion was so different across diet treatments, it is not possible
to interpret the effect of diet on PTH sensitivity of renal calcium reabsorption.

Because cows fed low-DCAD diets do excrete more Ca in their urine, it has been suggested that renal reabsorption of this potential pool of Ca (5–7 g of Ca/d) could provide the Ca needed to prevent development of hypocalcemia and milk fever. Schonewille et al. (1999) fed low-DCAD diets to cows and noted that they excreted about 6 g of Ca/d via the urine. When they then challenged the cow’s Ca homeostasis mechanisms by intravenous infusion with sodium EDTA to reduce blood ionized Ca and induce PTH secretion, they found that urinary Ca excretion declined to almost zero. Their data suggested that the extra 6 g of Ca normally excreted into the diet on a low-DCAD diet was returned to the blood and could help the cow maintain Ca homeostasis.

Our data do not replicate the near-complete reabsorption of Ca from urine found in the Schonewille et al. (1999) study. In the face of continued production of acidic urine, our low-DCAD cows were only able to reabsorb a small portion of the Ca in the glomerular filtrate when treated with PTH. Urine Ca:creatinine ratios did not decrease significantly in either diet group during the administration of PTH in our study (Figure 7). Our model did not impose a Ca challenge upon the cows as did the model of Schonewille et al. (1999), and it may be a mistake to dismiss the importance of urine Ca reabsorption as an aspect of low-DCAD diet mechanism of action in preventing milk fever.

Parathyroid hormone should increase the renal threshold for Mg excretion by the kidney. This should result in an increase in plasma Mg and a reduction in urine Mg:creatinine ratio as more of the Mg is reabsorbed from the glomerular filtrate (Goff, 1999). During the first 9 h of PTH treatment, plasma Mg increased in both groups of cows (Figure 2). The rate of increase in plasma Mg was greater in cows fed the low-DCAD diet during the first 9 h, but this may reflect the fact that their baseline plasma Mg was lower than that in the high-DCAD cows. Urine Mg:creatinine ratios decreased in both low- and high-DCAD cows upon PTH treatment at a similar rate despite differences in baseline (time 0) Mg:creatinine ratios (Figure 7).

Taken together, our data demonstrate that the high-DCAD diet induced a compensated metabolic alkalosis, which reduced the ability of the cows to respond to PTH. The diet induced a state of pseudohypoparathyroidism. Neither dietary Ca nor Mg differed across treatments, so neither can be considered a factor influencing tissue response to PTH in this study. Mayer et al. (1969) had already demonstrated that the parathyroid glands recognize the onset of hypocalcemia in milk fever cows and the parathyroid glands respond by greatly increasing PTH secretion. Martig and Mayer (1973) originally suggested that pseudohypoparathyroidism was the cause of milk fever because the response of late-gestation
cows to injections of parathyroid extracts was greatly decreased compared with that of mid-lactation cows. In human medicine, people diagnosed with pseudohypoparathyroidism have been identified as being resistant to the effects of PTH. They often have hypocalemia and very high circulating serum levels of PTH. The inability of these people to respond to PTH is often due to a genetic defect in the G-stimulatory protein coupled to the PTH receptor, which when properly activated by PTH, should result in intracellular accumulation of the second messenger, cyclic AMP. Our data support the observations of Martig and Mayer (1973) that transient pseudohypoparathyroidism is a major factor contributing to hypocalemia and milk fever. An argument can be made that testing tissue responsiveness several weeks before parturition does not reflect how the tissues will respond at the time of calving. Unfortunately, testing tissue responsiveness in periparturient cows is complicated by many factors. Goff et al. (1989) and Phillippo et al. (1994) saw diminished production of 1,25(OH)2D in the face of elevated PTH in periparturient cows fed alkalogenic high-DCAD diets. This suggests that the model we used is translatable to the periparturient cow and demonstrates that diet DCAD can influence response of tissues to PTH, and that blood pH is an integral part of the diet DCAD effect. The influence of acid-base status on calcium metabolism is not restricted to the ruminant. Beck and Webster (1976) observed that acute metabolic acidosis directly increases Ca mobilization from rat bone and augments the effect of PTH to mobilize calcium from bone. In their studies, acidosis directly inhibited renal tubular reabsorption of calcium, but augmented the effect of PTH to increase renal tubular reabsorption of calcium. Martin et al. (1980) found that the uptake of PTH by isolated perfused dog bone was enhanced under conditions of metabolic acidosis, resulting in greater release of cyclic AMP, the second messenger elicited when PTH binds to its receptor.

Acid-base status has been shown to affect responsiveness of tissues to other hormones as well. Under conditions of metabolic acidosis, hepatocytes fail to respond to growth hormone by producing insulin-like growth factor-I (Brüngger et al., 1997). Acidosis can also induce insulin resistance in tissues, complicating treatment of diabetes in humans, and may play a role in ketosis of cows (Bigner et al., 1996; Mitch, 2006). Metabolic acidosis alters the tertiary structure of the insulin receptor, reducing its affinity for insulin. It does not alter receptor number (Igarashi et al., 1993). In the case of the dairy cow, our data support that the condition of metabolic alkalosis induced by a high-DCAD diet results in a poor response of Ca regulating tissues to PTH resulting in acute hypocalemia, which is sometimes so severe as to result in milk fever. The typical dairy cow diet before calving is relatively high in K. This induces a metabolic alkalosis, causing a state of pseudohypoparathyroidism, and preventing rapid successful adaptation of the cow to the Ca demands of lactation. We speculate that the tertiary structure of the PTH receptor–G-protein complex is altered during metabolic alkalosis, reducing its affinity for PTH and resulting in pseudohypoparathyroidism. The loss of tissue sensitivity to PTH is not a total loss of ability to respond, as it is in humans with mutations in the PTH receptor–G-protein complex. In cattle, it appears to be a reduction in response, because very high levels of PTH over an extended period can eventually overcome the tissue resistance to PTH. For example, feeding very low Ca diets (less Ca absorbed than the cow requires) to stimulate endogenous PTH secretion in the weeks before calving and exogenous administration of pharmacologic doses of PTH for several days just before calving have both been successfully used to prevent milk fever, even when diet DCAD was high (Goings et al., 1974; Green et al., 1981; Goff et al., 1986). Low-DCAD diets induce a compensated metabolic acidosis that restores tissue sensitivity to PTH. This allows Ca homeostasis to proceed normally and returns blood Ca to normal quickly, preventing development of subclinical hypocalemia or milk fever.

REFERENCES


