Technical note: Comparison of 4 electronic handheld meters for diagnosing hyperketonemia in dairy cows

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ABSTRACT

The objective of this study was to evaluate 4 handheld ketone meters for use in on-farm β-hydroxybutyrate (BHB) monitoring of hyperketonemia in transition dairy cows. Blood samples taken from 250 Holstein cows between 262 d pregnant and 15 d in milk were evaluated on 4 different handheld ketone meters: Precision Xtra (Abbott Laboratories, Abbott Park, IL), TaiDoc (Pharmadoc, Lüdersdorf, Germany), Nova Max (Nova Biomedical, Billerica, MA), and Nova Vet (Nova Biomedical). Samples were screened using the Precision Xtra and tested on the remaining 3 m if the sample BHB concentration fell into predetermined ranges. A total of 89 samples were used for analysis. Performance of each meter was compared with the average of 2 plasma BHB concentrations both determined by a gold standard spectrophotometric Randox assay performed at 2 independent laboratories. Agreement between the 2 laboratories was very strong (Pearson correlation = 0.998). All meters had Pearson correlation coefficients greater than 0.95. The Precision Xtra and TaiDoc meters were 100.0% sensitive and 73.5% specific at a BHB concentration cut point of 1.2 mmol/L. The Nova Vet and Nova Max meters had sensitivities of 94.9 and 74.4% and specificities of 91.8 and 100.0%, respectively, at the same cut point. Agreement between the gold standard and the handheld meter was the best for the Nova Vet meter when evaluated using a Bland Altman graph with a mean BHB difference of 0.08 mmol/L. Trends in bias were noted with the Precision Xtra and Nova Max meters resulting in increasing average discrepancy between the gold standard and the handheld meter for both at higher plasma BHB concentrations and mean BHB differences of −0.34 and 0.26 mmol/L, respectively. The coefficient of variation was <10% for the Precision Xtra, TaiDoc, and Nova Vet meters, and <15% for the Nova Max meter. We conclude that the TaiDoc and Nova Vet meters, similar to the already validated Precision Xtra meter, are acceptable for use in on-farm testing for monitoring and treatment of hyperketonemia.

Key words: dairy cow, hyperketonemia, β-hydroxybutyrate, cowside meter

Technical Note

Most dairy cows undergo a period of negative energy balance (NEB) as they transition from late gestation to early lactation, a result of both an increase in energy requirements due to milk production and a decrease in DMI (Bauman and Currie, 1980; Baird, 1982; Herdt, 2000). In response to NEB, cows begin to break down fat stores; this increase in lipolysis releases fatty acids, which, among other avenues, leads to production of ketone bodies (Palmquist et al., 1969; Herdt, 2000). Thus, elevated concentrations of blood fatty acids and ketone bodies (e.g., BHB) are part of a normal adaptation of dairy cows to NEB in early lactation. However, excessive blood concentrations of fatty acids or BHB indicate a poor adaptation to NEB, which leads to an increased risk of detrimental health and production outcomes (Ospina et al., 2010; Chapinal et al., 2012; McArt et al., 2013). In addition, elevated levels of fatty acids and BHB can be detrimental to immune function (Hammon et al., 2006; Contreras et al., 2010; Ster et al., 2012). The incidence of elevated blood BHB ≥1.2 mmol/L in herds, or hyperketonemia (HYK), averages 40 to 60% in early lactation, and is thus a widespread issue in the dairy industry (Duffield et al., 1998; McArt et al., 2012).

Due to the high incidence of HYK, on-farm testing is important for monitoring and treatment of this disease. Recently, most on-farm testing has been performed using the Precision Xtra meter (Abbott Laboratories, Abbott Park, IL), a handheld blood ketone meter originally developed for human use. This meter has been well validated against the gold standard (spectrophotometric determination) for bovine BHB concentration in blood and found to be much more sensitive and specific
than previous handheld urine and milk tests at diagnosing HYK (Oetzel, 2004; Carrier et al., 2004; Iwersen et al., 2009).

Over the past few years, additional BHB meters have been developed both for human and veterinary use, including the TaiDoc meter (Pharmadoc, Lüdersdorf, Germany) and the Nova Max and Nova Vet meters (Nova Biomedical, Billerica, MA). These meters have not yet been rigorously evaluated for their accuracy. The objective of this study was to determine the diagnostic performance of these additional handheld meters for cowside use to provide the dairy industry with additional options for on-farm HYK diagnosis and monitoring.

We conducted 2 experiments using the Precision Xtra, TaiDoc, Nova Max, and Nova Vet meters. Experiment 1 was designed to determine each meter’s linearity compared with the gold standard method across a wide range of blood BHB concentrations. Experiment 2 was designed to determine each meter’s repeatability. Heparinized blood samples were collected from 4 dairy farms in New York State from October until December 2015. Whole blood was collected from 250 Holstein cows between 262 d pregnant and 15 DIM using 10-mL heparinized vacutainer tubes and 20-gauge, 2.54-cm blood collection needles. Cows were sampled weekly during the dry period and daily following calving resulting in approximately 1,400 samples. Blood samples remained at room temperature until testing for HYK using a Precision Xtra meter, which occurred within 4 h of collection. As previous reports have shown no difference between BHB concentrations determined with handheld meters in fresh whole blood and samples stored at room temperature or with various additives, all samples were treated as though they were fresh, whole blood (Gordon et al., 2013; Iwersen et al., 2013; Megahed et al., 2015). Prior to testing, each blood sample was inverted gently 5 times; samples were evaluated following manufacturer guidelines.

For experiment 1, the first 5 blood samples with BHB concentrations (determined using a Precision Xtra meter) at or within previously chosen concentrations or concentration ranges were enrolled for a goal of 100 total blood samples. β-Hydroxybutyrate concentrations ≤0.5 mmol/L were measured as one range, concentrations from 0.6 through 1.5 mmol/L were evaluated in 0.1 mmol/L increments, concentrations from 1.6 through 2.5 mmol/L were evaluated in 0.2 mmol/L increments, concentrations from 2.6 through 4.0 mmol/L were evaluated in 0.5 mmol/L increments, and concentrations >4.0 mmol/L were measured as one range. No more than 5 samples in each BHB concentration or concentration range were enrolled; however, some BHB concentrations or concentration ranges had fewer than 5 enrolled samples. The following concentrations or concentration ranges contained less than 5 samples: 1.5 mmol/L (n = 3), 2.2 to 2.3 mmol/L (n = 4), 2.4 to 2.5 mmol/L (n = 1), 2.6 to 3.0 mmol/L (n = 4), 3.1 to 3.5 mmol/L (n = 4), and 3.6 to 4.0 mmol/L (n = 2). A total of 89 samples were measured as one range, concentrations from 0.6 through 1.5 mmol/L were evaluated following manufacturer guidelines.

Following analysis, blood samples were centrifuged (10 min, 10,000 × g, 20°C) and plasma was stored in 2 aliquots at −80°C. After completion of meter testing, samples were submitted to the New York State Animal Health Diagnostic Center (AHDC, Ithaca, NY) and Immunochemical Assay Laboratory (Saalfeld, Germany) for BHB concentration determination using gold standard spectrophotometric Randox assays (Randox Laboratories Ltd., Crumlin, Co. Antrim, UK). For the 2 laboratories, intra-assay variation was less than 1% at both high and low control levels and the inter-assay variation was less than 5% at both control levels. Agreement between the AHDC and Immunochemical Assay Laboratory was determined using simple linear regression with JMP Pro (SAS Institute Inc., Cary, NC). Plasma BHB concentrations determined by the AHDC and Immunochemical Assay Laboratory had a very strong linear relationship (Pearson correlation = 0.998, slope = 1.03, P < 0.001). The gold standard for our analysis was calculated as the average BHB concentration between the AHDC and Immunochemical Assay Laboratory.

Linearity for each of the meters was determined using simple linear regression with JMP Pro, and test agreement between the gold standard and the handheld meter was determined via Bland Altman plots created by MedCalc (MedCalc Software, Ostend, Belgium). For repeated measurements, coefficients of variation were calculated for each meter in addition to the average variation from the mean. The sensitivity and specificity for each meter were determined at 1.2 and 3.0 mmol/L.

A total of 89 samples fit our criteria. One sample was excluded from analysis due to plasma
clotting during transport for gold standard analysis, and one value for the Precision Xtra was excluded from interpretation due to a reading above the reportable range of the meter (>8.0 mmol/L). Regression graphs of blood BHB concentrations determined by the meters compared with the gold standard are in Figure 1; Pearson correlation coefficients for the 4 handheld meters are summarized in Table 1. All Pearson correlation coefficients were greater than 0.95 indicating a strong linear relationship for all meters. The correlation coefficient for the Precision Xtra was similar to a previously reported correlation coefficient of 0.95 (Iwersen et al., 2009).

Performance of the 4 handheld meters for classification of a cow as hyperketonemic at blood BHB con-

Table 1. Pearson correlation coefficients for 4 handheld meters comparing blood BHB concentrations to plasma BHB concentrations determined by a gold standard spectrophotometric Randox assay ranging from 0.3 to 7.9 mmol/L (n = 89 samples from 64 Holstein cows between 274 d of gestation and 14 DIM)

<table>
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<tr>
<th>Test</th>
<th>Slope</th>
<th>r</th>
<th>P-value</th>
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<td>TaiDoc</td>
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<td>Nova Vet</td>
<td>0.87</td>
<td>0.97</td>
<td>&lt;0.001</td>
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1Randox (Randox Laboratories Ltd., Crumlin, Co. Antrim, UK).
2Precision Xtra (Abbott Laboratories, Abbott Park, IL), TaiDoc (Pharmadoc, Lüdersdorf, Germany), Nova Max (Nova Biomedical, Billerica, MA), and Nova Vet (Nova Biomedical).

Figure 1. Correlation of BHB concentrations determined in plasma by a gold standard spectrophotometric Randox assay (Randox Laboratories Ltd., Crumlin, Co. Antrim, UK) to whole-blood BHB concentrations determined with 4 handheld meters: (A) Precision Xtra (Abbott Laboratories, Abbott Park, IL), (B) TaiDoc (Pharmadoc, Lüdersdorf, Germany), (C) Nova Max (Nova Biomedical, Billerica, MA), and (D) Nova Vet (Nova Biomedical; n = 89 samples from 64 Holstein cows between 274 d of gestation and 14 DIM).
centrations of 1.2 and 3.0 mmol/L are summarized in Table 2. For the Precision Xtra meter, the sensitivity of 100.0% and the specificity of 73.5% was different than previous reports of 88 to 96% and 96 to 98%, respectively, at a cut point of 1.2 mmol/L (Iwersen et al., 2009; Voyvoda and Erdogan, 2010). This variation is most likely due to the range of BHB concentrations that we chose during data collection, favoring 1.0 to 1.4 mmol/L range, with a higher than normal percentage of samples chosen for BHB concentrations >1.4 mmol/L. This resulted in a nonrepresentative sample population clustered tightly around the cut point; a more representative population with a more uniform coverage across BHB concentrations would most likely result in a higher specificity. A similar finding and explanation can be made for the TaiDoc meter. Additionally, concerning the Precision Xtra meter, there is potential variation between batches of strips, which may have also resulted in some of the differences seen between studies.

The Nova Max meter had the lowest sensitivity (74.4%) and highest specificity (100.0%) of all meters at a 1.2 mmol/L cut point due to the consistently lower concentration reading compared with plasma BHB concentrations. Bovine blood has a lower hematocrit than humans, which is the most likely cause for the lower readings (Lane and Campbell, 1969; Kirk and Davis, 1970). It is important to note that when using the Nova Vet meter for bovine blood, Nova Biomedical recommends a 1.25 calibration slope factor to correct for this difference in hematocrit; this calibration cannot be performed on the Nova Max meter. The sensitivity and specificity of the Nova Vet meter without the calibration slope (i.e., directly out of the packing box) were 64.1 and 100.0%, respectively (data not shown), which is not acceptable for diagnostic use. The TaiDoc meter was designed for use with bovine blood. The Nova Vet meter, when calibrated appropriately, had a sensitivity and specificity of 94.9 and 91.8%, respectively, at a cut point of 1.2 mmol/L, which is most comparable to the Precision Xtra values previously reported by Iwersen et al. (2009) and Voyvoda and Erdogan (2010).

With few samples above 3.0 mmol/L, sensitivity and specificity at this cut point should be interpreted with caution. If we had more samples with BHB concentrations ranging from 2.0 to 4.0 mmol/L, we expect that, due to our determined bias of these meters, the specificity of the Precision Xtra and TaiDoc meters would decrease and the sensitivity of the Nova Max and Nova Vet meters would decrease. The sensitivity at this cut point was the highest for the Precision Xtra, TaiDoc, and Nova Vet meters and the poorest for the Nova Max meter. The specificity of all meters was acceptable.

Clinically, it can be argued that a test for HYK with a high sensitivity is more beneficial than a test with a high specificity, as the economic and health benefits of treating HYK positive cows outweigh the negative consequences of treating nonketotic animals. Although all meters were quite variable in their reported concentrations in samples with BHB >3.0 mmol/L, as treatment plans for severely ketotic animals are often unchanged at these higher concentrations (BHB ≥3.0 mmol/L; Oetzel, 2004), the clinical relevance of this variation is debatable.

Test agreement between plasma BHB concentrations, as determined by the gold standard, and whole blood BHB concentrations determined by the 4 difference handheld meters is shown in Figure 2. The Nova Vet had the least bias compared with plasma BHB concentrations with a mean difference of only 0.08 mmol/L and the most even distribution of points around the mean. Clinically, the bias seen with the Nova Vet meter at the 1.2 mmol/L cut point is negligible; a difference of 0.08 mmol/L would not affect classification of HYK. The TaiDoc, on average, read slightly high, with a mean BHB difference compared with plasma of −0.21 mmol/L. The Precision Xtra also read high with a mean difference of −0.34 mmol/L, and the mean dif-

<table>
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<td>Nova Vet</td>
<td>94.9</td>
<td>100.0</td>
<td>91.8</td>
<td>100.0</td>
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</tbody>
</table>

1Randox (Randox Laboratories Ltd., Crumlin, Co. Antrim, UK).
2Precision Xtra (Abbott Laboratories, Abbott Park, IL), TaiDoc (Pharmadoc, Lüdersdorf, Germany), Nova Max (Nova Biomedical, Billerica, MA), and Nova Vet (Nova Biomedical).
ference appeared to increase as the BHB concentration increased. For the Precision Xtra, TaiDoc, and Nova Max, the bias would result in a few additional animals being misclassified; however, in a herd monitoring scheme, these numbers would be minimal. Adjustment to a cut point of 1.4 mmol/L for the Precision Xtra and TaiDoc would also correct for some of the misclassification.

Within-meter variations in blood BHB concentration repeatedly measured 10 times at 3 different plasma BHB concentrations are summarized in Table 3. The 3 concentrations for repeated sampling were based on predetermined concentrations measured in whole blood with the Precision Xtra Meter of 0.6, 1.4, and 3.2 mmol/L; however, plasma BHB concentrations showed that repeated sampling actually occurred at 0.6, 1.1, and 2.3 mmol/L, respectively. All meters, aside from the Nova Max, showed a <10% coefficient of variation at all BHB evaluated concentrations. Average deviation for the mean BHB concentration for each meter was also extremely small (i.e., <0.05 mmol/L), which is clinically negligible. For humans, clinical standards for point-of-care glucose meters state that ≥95% of the values obtained with a meter should be within ±15% of each other (International Organization for Standardization, 2013). Using these standards, the repeatability of all meters is considered acceptable at all BHB evaluated concentrations.

In conclusion, Nova Vet and TaiDoc meters performed well and are acceptable alternatives to the Precision Xtra for on-farm BHB analysis. Additional research on a larger sample size with simple random sampling

Figure 2. Differences of BHB concentrations determined in plasma by a gold standard spectrophotometric Randox assay (Randox Laboratories Ltd., Crumlin, Co. Antrim, UK) and whole-blood BHB concentrations determined with 4 handheld meters: (A) Precision Xtra (Abbott Laboratories, Abbott Park, IL), (B) TaiDoc (Pharmadoc, Lüdersdorf, Germany), (C) Nova Max (Nova Biomedical, Billerica, MA), and (D) Nova Vet (Nova Biomedical; n = 89 samples from 64 Holstein cows between 274 d of gestation and 14 DIM).
would allow us to better understand the performance of these meters at higher BHB concentrations. The low sensitivity of the Nova Max meter and its marginal repeatability should exclude it from use as a tool for HYK testing in dairy cows.

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